Full Length Research Paper

Modelling the effect of supplementing elephant grass with lablab and desmodium on weight gain of dairy heifers under stall-feeding system

F. Tibayungwa¹*, J. Y. T. Mugisha² and M. Nabasirye³

¹Department of Animal Science, Makerere University, P. O. Box 7062, Kampala, Uganda. ²Department of Mathematics, Makerere University, P. O. Box 7062, Kampala, Uganda. ³Department of Crop Science, Makerere University, P. O. Box 7062, Kampala, Uganda.

Accepted 25 August, 2010

This study reports on a simulation growth model developed to predict daily gain (DG) of dairy cattle heifers. The model input parameters are gross energy (GE), ash, crude protein (CP), organic matter digestibility (OMD), dry matter (DM), protein degradation variables and heifer initial body weight. Results from the simulation model show that at low levels of CP in elephant grass (*Pennisetum purpureum*) there is improved DG when supplemented with desmodium (*Desmodium* spp.) or lablab (*Lablab purpureus*) but as the CP in elephant grass increases there is reduced benefit from supplementation and at CP \geq 100 g/kgDM there is no improvement in DG due supplementation. Supplementation with either lablab or desmodium at same percentage of diet had similar effect on DG. It is concluded that these two forage legumes could improve heifer growth, consequently reducing the time from weaning to mating weight of heifers in smallholder dairying where the CP content of elephant grass is low.

Key words: Elephant grass, daily gain, supplement, desmodium, lablab.

INTRODUCTION

Stall-feeding of dairy cattle was introduced and promoted in Uganda by non-governmental organizations (NGO) and the Uganda government to improve household nutrition, incomes and food security among resource poor households. A study by MAAIF(1996) found that stallfeeding system had the highest economic returns compared to other cattle management systems. However, reproductive performance reduce the profitability of smallholder dairy farmers Nakiganda et al. (2006). For example, the age at first calving in stall-fed dairy cattle, is 2.5 years (Bebe et al., 2003; Twinamatsiko, 2001) as compared to 2 years in developed world and 28 months recommended for smallholder (MLD, 1991).

In Uganda where smallholder dairying is primarily dependent on elephant grass as the sole source of feed, replacement heifers take long to attain mating weights. Typically for ruminants on forage, energy and then protein are the primary limiting nutritional requirements (Freer et al., 1997), but elephant grass has been found to meet most of the energy requirements in smallholder dairying (Muia et al., 2000a).

According to ARC (1984) content of a diet should be \geq 120 g/kgDM if moderate production in dairy cattle is to be attained. Yet CP in elephant grass has been found to decline from 200 - 50 g/kgDM (Muia et al., 1999) and 200 - 50 g/kgDM (Ogwang and Mugerwa, 1976), from 3 - 15 weeks and 6 - 12 weeks of age respectively. One way to improve the low CP diet is to feed forage legumes. In Kariuki et al. (1999), they reported a significant increase in weight gain of heifers fed elephant grass supplemented with forage legumes. The two most popular forage legumes in Uganda are lablab and desmodium

^{*}Corresponding author. E-mail: ftibayungwa@agric.mak.ac.ug. Tel: +256-414-532269. Fax: +256-414-531641.

Abbreviation: DG, Daily gain; GE, gross energy; CP, crude protein; OMD, organic matter digestibility; DM, dry matter.

| Age | DM | Ash ^b | СР | GE | ADIN | IVDMD | Reference |
|-----|-------|------------------|-------|------|------|-------|---------------------------|
| - | 192 | 159 | - | - | - | 808 | Hassan et al. (1983) |
| - | 234 | 152.8 | 196.9 | | - | - | Hassan et al. (1979) |
| 10 | 182.7 | - | 81.8 | 15.4 | - | - | Muia et al. (2001a) |
| 15 | 238.6 | - | 53.3 | 16.9 | - | - | Muia et al. (2001a) |
| 10 | 180 | - | 83 | 16 | - | - | Muia et al. (2001b) |
| 15 | 240 | - | 53 | 17 | - | - | Muia et al. (2001b) |
| 10 | 183.1 | - | 84.1 | 16.1 | - | - | Muia et al. (2000) |
| 15 | 237.8 | - | 53 | 16.8 | - | - | Muia et al. (2000) |
| - | 176 | - | 68.4 | - | - | - | Nyambati et al. (2003) |
| - | - | 111 | 115.4 | - | 1.3 | - | Kabi et al. (2005) |
| Mc | - | 136 | 61 | 15.6 | - | 560 | Mlay et al. (2006) |
| - | - | - | 111.5 | - | - | 638.4 | Ogwang and Mugerwa (1976) |
| 6-8 | - | 124.9 | 102.5 | - | - | 692 | Mpairwe et al. (1998) |

Table 1. Nutrient composition and digestibility, energy and age of Napier grass^a.

^a DM, Ash, CP, ADIN, IVDMD in g/kgDM; GE in MJ/kgDM; Age in weeks. ^b Where DOMD is not known, then Ash is used according to Equation 2). ^c M refers to mature, no specific age given.

Table 2. Chemical composition, energy and crude protein degradation of lablab^a.

| DM | СР | OMD | Ash ^b | GE | ADIN | а | b | С | Reference |
|-----|-------|-----|------------------|-------|------|--------|--------|-------|-----------------------------|
| - | 170 | 560 | - | - | - | - | - | - | Murphy and Colucci (1999) |
| - | 157 | - | - | - | - | 0.237 | 0.691 | 0.105 | Mpairwe et al. (2003a) |
| - | | - | - | - | - | 0.244 | 0.676 | 0.153 | Mpairwe et al. (2003b) |
| - | 158 | 597 | - | - | - | - | - | - | Kabirizi (2006) |
| - | 163 | - | - | - | - | - | - | - | Nyambati et al. (2003) |
| - | 180 | - | 119.4 | - | - | 0.2479 | 0.6363 | 0.14 | Melaku et al. (2003) |
| 215 | 198 | - | 115.0 | - | - | - | - | - | Linga et al. (2003) |
| - | 254 | - | 114.0 | - | 2.44 | - | - | - | Mupangwa et al. (2006) |
| - | 216 | - | 114.0 | - | 0.71 | - | - | - | Mupangwa et al. (2006) |
| 170 | 174 | - | - | - | - | - | - | - | Mbuthia and Gachuiri (2003) |
| - | 191.8 | - | - | 13.16 | - | - | - | - | Nworgu and Ajayi (2005) |
| - | 128 | - | 129 | - | - | - | - | - | Osuhor et al (2004) |
| - | - | - | - | - | 1.26 | - | - | - | Mupangwa et al. (2003) |

^a DM, Ash, CP, ADIN in g/kgDM; GE in MJ/kgDM; ^b Where DOMD is not known, then ash is used according to Equation (2).

because they are easily intercropped with elephant grass and food crops. However, it is not yet established at what CP level in elephant grass these legumes should be included or whether they have different effect on heifer DG.

The objective of this research was to study the effect of supplementing elephant grass at different levels of CP with lablab and desmodium on DG in dairy cattle heifers, by extending a simulation model of heifer growth developed in Tibayungwa et al. (2009), in a stall-feeding dairy system.

MATERIALS AND METHODS

This section summarises the procedures, assumptions and

equations used to develop a growth model of dairy heifers from weaning to mating weight, for elephant grass supplemented with lablab and desmodium in a stall-feeding smallholder dairying system.

Feed composition

Feed parameters of elephant grass, lablab and desmodium are given in Tables 1, 2 and 3 respectively.

Energy value of feed

The energy value of feed was estimated based on AFRC (1993) as follows:

 $ME(MJ/kgDM) = 0.0157 \times DOMD(g/kgDM)$ (1)

| Species | DM | СР | OMD | Ash | GE | ADIN | а | b | С | Reference |
|-------------|-----|-------|-------|------|-------|------|-------|-------|--------|-------------------------|
| D.intortum | - | 120 | | 87 | | | | | | Nurfeta et al. (2008) |
| D.uncinatum | - | | | 84.3 | | | 0.311 | 0.414 | 0.065 | Baloyi et al. (2008) |
| D.uncinatum | 270 | | 496 | 40 | | | | | | Milford (1967) |
| D.intortum | 486 | 105 | | | 15.0 | | | | | Aregheore et al. (2006) |
| D.intortum | - | 118.1 | | 55.1 | | | 0.214 | 0.216 | 0.0 2 | Mghen et al. (1996) |
| D.ucinatum | - | 163.1 | | 85.1 | | | 0.291 | 0.423 | 0.0 24 | Mghen et al. (1996) |
| D.intortum | - | 229.4 | | | 20.47 | | | | | Stobbs (1971) |
| D.intortum | - | 199 | | 98 | | | | | | Getachew et al. (2000) |
| D.uncinatum | - | 134.5 | 642.9 | 104 | | 2.6 | | | | Jingura et al. (2001) |

Table 3. Nutrient composition and digestibility, degradability, energy and age of Desmodium spp.^a

^aDM = Dry matter (g/kg); DOMD = digestible organic matter (g/kgDM); CP = crude protein (g/kgDM); ADIN = acid detergent insoluble nitrogen (g/kgDM); GE = gross energy (MJ/kgDM); a = water soluble fraction; b = potentially degradable nitrogen other than water soluble fraction; c = degradation rate per hour of the *b* fraction.

(**a**)

where ME is metabolisable energy; DOMD is Digestible Organic Matter in a feed, and is estimated as

 $DOMD = OMD \times (1000 - total ash) / 1000$ (2)

where OMD is Organic Matter Digestibility (g/kg)

$$FME(MJ / kgDM) = ME \times (0.467 + 0.00136 \times ODM - 0.00000115 \times ODM^{2})$$
(3)

where FME (MJ/kgDM) is fermentable metabolisable energy; ODM is Oven Dry Matter content (g/kg)

Protein value of feed

Estimation of the metabolisable Protein (MP) from Crude Protein (CP) involves the following calculations (AFRC, 1993) (definitions of symbols used are in Table 4):

$$UDP = CP - \{QDP + SDP\}$$
⁽⁴⁾

$$SDP = \left\{ \left(b \times c \right) / \left(c + r \right) \right\} \times CP \tag{5}$$

$$QDP = a \times CP$$
 (6)

where r is calculated as follows

$$r = -0.024 + 0.179 \left\{ 1 - e^{(-0.278L)} \right\}$$
(7)

where L is level of feeding as a multiple of MJ of ME for maintenance.

$$MCP = FME \times y \tag{8}$$

where y is microbial protein yield in the rumen (gMCP/MJ of FME), and is calculated as

$$y = 7.0 + 6.0 \left\{ 1 - e^{(-0.35L)} \right\}$$
(9)

$$D U P = 0.9 \{ U D P - 6.25 A D I N \}$$
(10)

$$DMTP = 0.6375MCP \tag{11}$$

$$MP(g / d) = 0.6375MCP + DUP$$
(12)

$$ERDP = 0.8QDP + SDP \tag{13}$$

If ERDP supply is less than (or equal to) ERDP required, then

$$MCP(g/d) = ERDP(g/d)$$
(14)

Else

$$MCP(g/d) = FME(MJ/d) \times y(gMCP/MJFME)$$
(15)

Estimation of intake

According to AFRC (1993) the dry matter intake (DMI) is estimated as follows:

$$DMI(kg / d) = MER / (M / D)$$
(16)

where MER is Metabolisable Energy Requirement (MJ/d), M/D is metabolisable energy (MJ/kgDM).

This estimation of DMI is appropriate where daily gain is predetermined and forage is available in adequate amount. In a case where the DMI depends on forage availability and daily gain is not known forehand, the intake can be estimated based on experimental observations. We used an estimate of 2.7% of body weight based on Kariuki et al. (1998) value of 2.94%, Diaz-Solis et al. (2006) value of 2.54% and Blomquist (2005) value of 2.5 - 3.0% of the body weight. Therefore

$$IF \quad Fa \ge 0.027 * W, \quad THEN \quad DMI = 0.027 * W,$$
$$ELSE \quad DMI = Fa \tag{17}$$

where Fa is available forage.

Protein requirements

The metabolisable protein is based on AFRC (1993). Metabolizable protein requirement for maintenance (kg/d) is estimated as

$$MP_m = 2.30W^{0.75}$$
(18)

Metabolizable Protein requirement for growth (kg/d) is estimated as

$$MP_{f} = C6 \{ 168.07 - 0.16869W + 0.0001633W^{2} \} \times \{ 1.12 - 0.1223\Delta W \} \times 1.695\Delta W$$
(19)

where MP_f is metabolizable protein requirement for liveweight gain (g/d), C6 is a correction factor ranging from 0.8 -- 1.0, W is liveweight of the animal (kg).

Energy requirements

The energy requirement is based on AFRC (1993) and is calculated as follows:

$$M_{mp}(MJ/d) = (E_m/k) \times \ln\{B/(B-R-1)\}$$
 (20)

where M_{mp} is ME requirement for both maintenance and production, E_m (MJ/d) is the sum of animal's fasting metabolism (F) and activity allowance (A = 0.0071 W) for zero-grazed heifers, R is the scaled energy retention.

The fasting metabolism, MJ/(kg fasted weight)^{0.67}, is defined as:

$$F = 0.53 \left(W \ / \ 1.08 \right)^{0.67} \tag{21}$$

The factors B and k are calculated from the efficiencies of utilization of ME as follows:

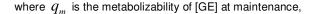
$$B = \frac{k_m}{\left(k_m - k_f\right)} \tag{22}$$

$$k = k_m \times \ln\left(k_m / k_f\right) \tag{23}$$

where k is the efficiency of utilization of ME (Metabolizable Energy) for a given metabolic process, B is a derived parameter to predict energy retention, k_m is the efficiency of utilization of ME for maintenance, k_f is the efficiency of utilization of ME for weight gain. Both k_m and k_f can be calculated as follows:

$$k_m = 0.35q_m + 0.503 \tag{24}$$

$$k_f = 0.78q_m + 0.006 \tag{25}$$



[ME]/[GE], where GE is the gross energy of a diet (MJ/d or MJ/kg DM).

Scaled energy retention (R) is calculated from:

$$E_{f} = C 4 \left(E V_{g} \times \Delta W \right)$$
⁽²⁶⁾

where C4 is the correction factor for metabolizable energy for heifers (= 1.1) and then:

$$R = \frac{E_f}{E_m}$$
(27)

where E_f is Net Energy retained in growing animal (MJ/d), E_m is Net Energy for maintenance (MJ/d).

Predicting live weight gain

Predicting live weight gain involves the following steps:

Step 1. Energy value of weight gain

This is given by the expression

$$E V_{s} = \frac{C 2 \left(4.1 + 0.0332W - 0.000009W^{2}\right)}{\left(1 - C 3 \times 0.1475\Delta W\right)}$$
(28)

where EV_g is energy value of tissue gained (MJ/kg), ΔW is liveweight change (kg/d), C2 is a correction factor (range 1.00 - 1.30) for mature body size and sex of animal; C3 is a correction factor for plane of nutrition (L), 1 when L > 1 and 0 when L < 1. These correction factors are given in AFRC (1993).

Step 2. Energy retention

Scaled energy retention (R) is as defined in Equation (27).

Step 3. Metabolisable Protein requirement for growth

Equation (19) is rearranged to estimate weight gain based on MP_f.

Step 4. Weight gain

Equation (26) is rearranged to give:

$$\Delta W = \frac{E_f}{\left(C \, 4 \times E \, V_e\right)} \tag{29}$$

By combining the two (28) and (29) Equations and that contain the term ΔW , we get:

$$\Delta W = \frac{E_f}{\left(C4X + 0.1475E_f\right)} \tag{30}$$

| Symbol | Definition | Units | |
|--------|---|---------------|--|
| а | Proportion of water soluble Nitrogen in the total Nitrogen of a feed | Unit-less | |
| ADIN | Acid detergent insoluble nitrogen in a feed | g/kgDM | |
| b | Proportion of potentially degradable N other than water soluble N of a feed | Unit-less | |
| С | Fractional rumen degradation rate per hour of the <i>b</i> fraction of feed N | Unit-less | |
| CP | Crude protein in of a diet or in a feed | g/kgDM, g/d | |
| DMTP | Digestible microbial true protein (= metabolizable protein from microbes) | g/d, g/kgDM | |
| DUP | Digestible undegraded protein (N x 6.25) | g/kgDM, g/d | |
| FME | Fermentable metabolizable energy of a diet | MJ/d, MJ/kgDM | |
| MCP | Microbial crude protein supply | g/d, g/kg | |
| MP | Metabolizable protein | g/d, g/kgDM | |
| MTP | Microbial true protein | g/d, g/kg | |
| QDP | Quickly degradable protein (N x 6.25) of a diet or in a feed | g/d, g/kgDM) | |
| r | Rumen digesta fractional outflow rate per hour | Unit-less | |
| SDP | Slowly degradable protein (N x 6.25) of a diet or in a feed | g/d, g/kgDM | |
| UDP | Undegradable dietary protein (N x 6.25) of a diet or in a feed | g/kgDM | |

Table 4. The definition of symbols and terminology.

where $X = C2(4.1+0.0332W - 0.000009W^2)$ is taken from

Equation 28

DESCRIPTION OF THE SIMULATION MODEL

It is assumed that the animal is not constrained in any other way except the supply of crude protein. It is further assumed that there are no inhibitory nor synergetic tendencies between the different forages used. The feed input parameters are DM, GE, ash, CP, CP degradation variables (*a*, *b*, *c*, see Table 4 for definitions), acid detergent insoluble nitrogen (ADIN). Animal input parameters are initial weight and level of feeding. The dry matter intake without supplementation is set at 2.7% of animal's weight as explained in section 2.4. The dry matter intake of elephant grass supplemented with forage legumes can increase by about 16.7% as reported in Kariuki et al. (1999); in the current study we used this estimate to raise the intake to 3.2% of body weight. All other parameters are calculated by the model using the respective coefficients as indicated in the equations. The microbial crude protein yield (*y*) is

determined by the amount of fermentable metabolisable energy (FME), If effective rumen degradable protein (ERDP) supply is less than (or equal to) ERDP required, then MCP = ERDP else MCP = FME multiplied by y.

After part of ME and MP have been used for maintenance, daily gain (DG) is dependent of the balance between Metabolisable Energy for growth (MEg) and Metabolisable Protein for growth (MPg); if potential growth due to metabolisable protein (Gp) is greater than the potential growth due to metabolisable energy (Ge), then MEg is considered limiting and the growth is determined by Ge. If potential growth due to metabolisable protein (Gp) is less than potential growth due to metabolisable energy (Ge), then MPg is considered limiting and the growth is determined by Gp. The simulated DG for the two forages is added to get the total DG which is then added to the weight to get a new liveweight (LW) and the process is repeated for the desired number of days. Since forages differ in nitrogen degradability, protein intakes were treated separately rather than summing them. The simulation model is coded in VENSIM 5.5 (The Ventana Simulation Environment, Ventana systems, Inc.), based on differential equations with a 1-day

time step.

Evaluation of the simulation model

The performance of the simulation model was evaluated by comparing model predictions to field data reported in Tables 1, 2 and 3 that were never used in the development of the model. The daily gain predicted on the basis of forage composition and animal weight and requirements were compared to the values reported in Kariuki et al. (1999).

RESULTS AND DISCUSSION

Model calibration and evaluation

Parameter selection for model calibration was based on the sensitivity of the parameter values in Tables 1, 2 and 3. The experiments from which these datasets were generated were either on the effect of supplementation on degradability or effect of supplementation on weight gain. Degradability parameters required as inputs for the simulation model were obtained from experiments that fall in the degradation category. Parameters that describe protein degradation in the rumen (a, b, c) and ADIN which contributes directly to fecal N levels, are highly variable (Webster, 1993) even when determined for the same samples at different laboratories. Therefore these parameters were selected as the starting point for the calibration. However, in cases where there was lack of data, the calibration datasets are the same ones used to derive parameters for the model, since they provide an indication of the models' performannce following manipulation of model parameters to improve accuracy (Hill et al., 2006). The simulation model predicted DG of 0.43 kg/day when heifers weighing 181 kg are fed elephant

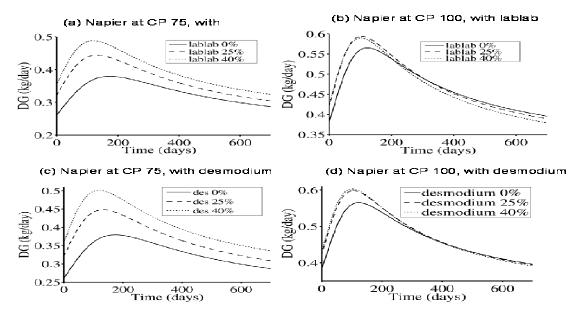


Figure 1. Daily gain when elephant grass is supplemented with Lablab and Desmodium at different levels. (a) Napier at CP 75 g/kgDM supplemented with lablab at different levels (b) Napier at CP 100 g/kgDM supplemented with lablab at different levels; (c) Napier at CP 75 g/kgDM supplemented with desmodium at different levels (d) Napier at CP 100 g/kgDM supplemented with desmodium at different levels.

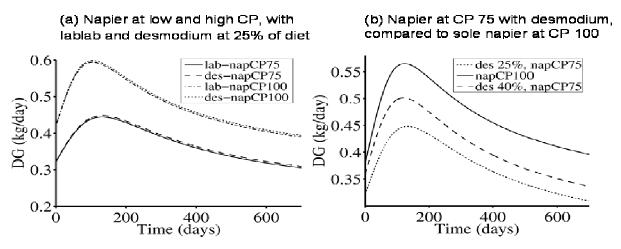


Figure 2. Daily gain of heifers when fed with elephant grass at two levels of CP and supplemented with Lablab and Desmodium. (a) Napier at CP 75 and 100 g/kgDM, supplemented with lablab and desmodium at 25% of diet; (b) Napier at CP 100 g/kgDM, compared to napier at CP 75% supplemented with desmodium at 25 and 40% of diet.

grass supplemented with desmodium for 120 days. This result is similar to field results of 0.42 kg/day reported by Kariuki et al. (1999).

Model use

According to Leng (1990), forages are considered as low quality if they have less than 80 g of CP/kgDM and high quality if 100 g of CP/kgDM and above. It is on this basis that CP 75 and 100 g/kgDM were chosen for model use. Figure 1 shows the DG of heifers fed elephant grass supplemented with lablab and desmodium. DG improved as the level of supplement increased (Figures 1a and c). However, at high levels of Napier CP (100 g/kgDM) the benefit of supplementation becomes less significant (Figures 1b and d), whereas at low napier CP (75 g/kgDM) supplementation up to 40% yielded less DG compared to unsupplemented napier at high CP (100 g/kgDM) as shown in Figure 1b. DG as a result of supplementation was similar for both lablab and desmodium (Figure 2a). Table 5 shows DG and time from weaning to

| | %Desmo | odium in diet | %Lablab in diet | | | |
|-----|-----------|---------------|-----------------|-----------|-----------|-----------|
| CP⁵ | 0 | 25 | 40 | 0 | 25 | 40 |
| 75 | 691(0.33) | 597(0.39) | 533(0.43) | 691(0.33) | 604(0.38) | 551(0.42) |
| 100 | 461(0.50) | 442(0.52) | 441(0.52) | 461(0.50) | 446(0.52) | 453(0.51) |

Table 5. DG and days from weaning to 300 kg LW of heifers fed napier grass supplemented with lablab and desmodium^a.

^aValues in parentheses are the DG in kg/day. ^bCP in Napier, g/kgDM.

mating weight of 300 kg LW of heifers fed napier grass supplemented with lablab and desmodium.

According to the MLD (1991) recommendations in the smallholder dairy farming systems, the weaning weight for dairy heifers is 70 kg and a target of 300 kg to be attained by 18 months of age for first service. For this target to be met the heifers are assumed to gain at least 0.5 kg/day. From the results of this study (Table 5) this target DG is not possible on low quality napier grass. Forages are considered as low quality if they have less than 80 g of CP/kgDM (Leng, 1990). As seen from Table 5 low quality elephant grass even when supplemented up to 40% of the diet, it is not possible to attain DG of 0.5 kg/day. However, with high quality elephant grass DG of 0.5 kg/day is possible with or without supplementation (Table 5). The lack of improvement in DG as a result of supplementation was also reported by Kariuki et al. (1999) when elephant grass was supplemented with desmodium; this is because of the high nutritive value of the elephant grass used. Therefore supplementing poor quality elephant grass with forage legumes improves DG but not necessarily so when the nutritive value of elephant grass is high. Although the simulation model predictions were similar to observed values of Twinamatsiko (2001) and Kariuki et al. (1999), the model has limitations in that only the averages for the observed values were available for model development and evaluation. But to optimize growth there is need to know the growth curve so that the appropriate amount of supplement is given at the right time. Therefore further research on heifer performance in smallholder dairy is needed to accumulate adequate data for developing and evaluating the simulation models of dairy heifer growth.

The simulation model indicates that lablab and desmodium have the same effect on DG and if they are to be of value they should be used when the elephant grass is poor quality, usually at an advanced stage of growth when it is low in CP. These two forage legumes could therefore improve heifer growth, consequently reducing the time from weaning to mating weight of heifers in smallholder dairying where the CP content of elephant grass is low.

REFERENCES

AFRC (1993). Energy and Protein Requirements of Ruminants. Aricultural and Food Research Council. CAB International, Wallingford, UK. ARC (1984). The nutrient requirement of ruminant livestock, Supplement No. 1. CAB, Slough, England.

- Aregheore EM, steglar TA, Ng'ambi JW (2006). Nutrient characterisation and in vitro digestibility of grass and legume/browse species-based diets for beef cattle in vanuatu. South Pacic J. Nat. Sci., 24(1): 20-27.
- Baloyi JJ, Ngongoni NT, Hamudikuwanda H (2008). Chemical composition and ruminal degradability of cowpea and silverleaf desmodium forage legumes harvested at different stages of maturity. Trop. Subtrop. Agroecosyst., 8: 81-91.
- Bebe BO, Udo HMJ, Rowlands GJ, Thorpe W (2003). Smallholder dairy systems in the Kenya highlands: cattle population dynamics under increasing intensification. Livest. Prod. Sci., 82(2-3): 211-221.
- Blomquist N (2005). How much will my cow eat? Frequently asked questions. Tech. rep., Alberta Ag-Centre, Alberta Agriculture, Food and Rural Development, Government of Alberta.
- Diaz-Solis H, Kothmann MM, Grant WE, De Luna-Villarreal R (2006). Use of irrigated pastures in semi-arid grazinglands: A dynamic model for stocking rate decisions. Agric. Syst., 88: 316-331.
- Freer M, Moore AD, Donnelly JR (1997). GRAZPLAN: decision support systems for Australian grazing enterprises. II The animal biology model for feed intake, production and reproduction and the GrazFeed DSS. Agric. Syst., 54: 77-126.
- Getachew G, Makkar HPS, Becker K (2000). Effect of polyethylene glycol on *in vitro* degradability of nitrogen and microbial protein synthesis from tannin-rich browse and herbaceous legumes. Br. J. Nutri., 84: 73-83.
- Hassan NI, Abdelaziz HM, El Tabbah AE (1979). Evaluation of some forages introduced to newly reclaimed areas in Egypt. World Rev. Anim. Prod. 15(2): 31-35.
- Hassan NI, Osman FA, Rammah AM (1983). Morphological characters, chemical composition and in vitro dry matter disappearance of new varieties of napier grass grown in Egypt. World Rev. Anim. Prod., 19(4): 35-40.
- Hill JO, Robertson MJ, Pengelly BC, Whitbread AM, Hall CA (2006). Simulation modelling of lablab (*Lablab purpureus*) pastures in northern Australia. Aust. J. Agric. Res., 57(4): 389-401.
- Jingura RM, Sibanda S, Hamudikuwanda H (2001). Yield and nutritive value of tropical forage legumes grown in semi-arid parts of zimbabwe. Trop. Grasslands, 35: 168-174.

Kabi F, Bareeba FB, Havrevoll O, Mpofu IDT (2005). Evaluation of

protein degradation characteristics and metabolisable protein of elephant grass (*Pennisetum purpureum*) and locally available protein supplements. Livest. Prod. Sci., 95 (1-2): 143-153.

- Kabirizi JM (2006). Effect of intergrating forage legumes in smallholder dairy farming systems on feed availability and animal performance. Ph.D. thesis, Makerere University, Kampala, Uganda.
- Kariuki JN, Gachuiri CK, Gitau GK, Tamminga S, van Bruchem J, Muia JMK, Irungu KRG (1998). Effect of feeding napier grass, lucerne and sweet potato vines as sole diets to dairy heifers on nutrient intake, weight gain and rumen degradation. Livest. Prod. Sci., 55(1): 13-20.
- Kariuki JN, Gitau GK, Gachuiri CK, Tamminga S, Muia JMK (1999). Effect of supplementing napier grass with desmodium and lucerne on DM, CP and NDF intake and weight gains in dairy heifers. Livest. Prod. Sci., 60(1): 81-88.
- Leng RA (1990). Factors affecting the utilisation of of 'poor quality' forages by ruminants particularly under tropical conditions. Nutr. Res. Rev., 3: 277-303.
- Linga SS, Lukefahr SD, Lukefahr MJ (2003). Feeding of *Lablab* purpureus forage with molasses blocks or sugar cane stalks to rabbit

fryers in subtropical south Texas. Livest. Prod. Sci., 80(3): 201-209.

- MAAIF (1996). Study Report on The Comparative Analysis of Cattle Management systems in Different Areas of Uganda. Ministry of Agriculture Animal Industry and Fisheries. Entebbe, Uganda.
- Mbuthia EW, Gachuiri CK (2003). Effect of inclusion of Mucuna pruriens and Dolichos lablab forage in napier grass silage on silage quality and on voluntary intake and digestibility in sheep. Trop. Subtrop. Agroecosyst., 1: 123-128.
- Melaku S, Peters KJ, Tegegne A (2003). In vitro and in situ evaluation of selected multipurpose trees, wheat bran and Lablab purpureus as potential feed supplements to tef (Eragrostis tef) straw. Anim. Feed Sci. Technol., 108(1-4): 159-179.
- Mghen DM, Hvelplund T, Weisbjerg MR (1996). Rumen degradability of dry matter and protein in tropical grass and legume forages and their protein values expressed in the AAT-PBV protein evaluation system.
 In: Ndikumana J, de Leeuw P (eds) Sustainable Feed Production and Utilization of Smallholder Livestock Enterprises in Sub-Saharan Africa. Proceedings of the Second African Feed Resources Network (AFRNET), Harare, Zimbabwe, 6-10 December 1993. AFRNET (African Feed Resources Network), Nairobi, Kenya.
- Milford R (1967). Nutritive value and chemical composition of seven tropical legumes and lucerne grown in subtropical south-eastern queensland. Aust. J. Exp. Agric. Anim. Husbandry, 7: 540-545.
- Mlay PS, Pereka A, Chikula PE, Balthazary S, Igusti J, Hvelplund T, Weisbjerg MR, Madsen J (2006). Feed value of selected tropical grasses, legumes and concentrates. VETERINARSKI ARHIV, 76(1): 53-63.
- MLD (1991). Zero Grazing: A Handbook on technical aspects. Ministry of Livestock Development. Nairobi, Kenya.
- Mpairwe DR, Sabiiti EN, Mugerwa JS (1998). Effect of dried Gliricidia sepium leaf supplement on feed intake, digestibility and nitrogen retention in sheep fed dried KW4 elephant grass (*Pennisetum purpureum*) ad libitum. Agrofor. Syst., 41: 139-150.
- Mpairwe DR, Sabiiti EN, Ummuna NN, Tegegne A, Osuji P (2003a). Integration of forage legumes with cereal crops. I. Effects of supplementation with graded levels of lablab hay on voluntary food intake, digestibility, milk yield and milk composition of crossbred cows fed maize-lablab stover or oats-vetch hay ad libitum. Livest. Prod. Sci., 79(2-3): 193-212.
- Mpairwe DR, Sabiiti EN, Ummuna NN, Tegegne A, Osuji P (2003b). Integration of forage legumes with cereal crops: II. Effect of supplementation with lablab hay and incremental levels of wheat bran on voluntary food intake, digestibility, milk yield and milk composition of crossbred cows fed maize-lablab stover or oats-vetch hay ad libitum. Livest. Prod. Sci., 79(2-3): 213{226.
- Muia JMK, Tamminga S, Mbugua PN (2000a). Effect of supplementing napier grass (*Pennisetum purpureum*) with sunfower meal or poultry litter-based concentrates on feed intake, live-weight changes and economics of milk production in Friesian cows. Livest. Prod. Sci. 67(1-2): 89-99.
- Muia JMK, Tamminga S, Mbugua PN, Kariuki JN (1999). Optimal stage of maturity for feeding napier grass (*Pennisetum purpureum*) to dairy cows in kenya. Trop. Grasslands, 33: 182-190.
- Muia JMK, Tamminga S, Mbugua PN, Kariuki JN (2000b). The nutritive value of napier grass (Pennisetum purpureum) and its potential for milk production with or without supplementation: a review. Trop. Sci., 40: 1-23.
- Muia JMK, Tamminga S, Mbugua PN, Kariuki JN (2001a). Effect of supplementing napier grass (*Pennisetum purpureum*) with poultry litter and sunflower meal based concentrates on feed intake and rumen fermentation in Friesian steers. Animal Feed Sci. Technol., 92(1-2): 113-126.

- Muia JMK, Tamminga S, Mbugua PN, Kariuki JN (2001b). Rumen degradation and estimation of microbial protein yield and intestinal digestion of napier grass (*Pennisetum purpureum*) and various concentrates. Anim. Feed Sci. Technol., 93 (3-4): 177-192.
- Mupangwa JF, Ngongoni NT, Hamudikuwanda H (2006). The effect of stage of growth and method of drying fresh herbage on chemical composition of three tropical herbaceous forage legumes. Trop. Subtrop. Agroecosyst., 6: 23-30.
- Mupangwa JF, Ngongoni NT, Topps JH, Acamovic T, Hamudikuwanda H (2003). Rumen degradability and post-ruminal digestion of dry matter, nitrogen and amino acids in three tropical forage legumes estimated by the mobile nylon bag technique. Livest. Prod. Sci., 79(1): 37-46.
- Murphy AM, Colucci PE (1999). A tropical forage solution to poor quality ruminant diets: A review of (*Lablab purpureus*). Livestock Res. Rural Dev., 11 (2).
- Nakiganda A, Mcleod A, Bua A, Phipps R, Upton M, Taylor N (2006). Farmers' constraints, objectives and achievements in smallholder dairy systems in Uganda. Livest. Res. Rural Dev., 18 (Article No.69).
- Nurfeta A, Tolera A, Eik LO, Sundstl F (2008). The supplementary value of different parts of enset (ensete ventricosum) to sheep fed wheat straw and desmodium intortum hay. Livestock Sci., 119 (1-3): 22-30.
- Nworgu FC, Ajayi FT (2005). Biomass, dry matter yield, proximate and mineral composition of forage legumes grown as early dry season feeds. Livest. Res. Rural Dev., 17(11).
- Nyambati EM, Sollenberger LE, Kunkle WE (2003). Feed intake and lactation performance of dairy cows offered napiergrass supplemented with legume hay. Livest. Prod. Sci., 83(2-3): 179-189.
- Ogwang BH, Mugerwa JS (1976). Yield response to N application and in vitro dry matter digestibility of elephant grass x bulrush millet hybrids. East Afri. Agric. For. J., 41: 231-242.
- Osuhor CU, Tanko RJ, Dung DD, Muhammad IR, Odunze AC (2004). Water Consumption of Yankasa Rams Fed a Basal Diet of Maize Stover-lablab Mixture. Pakistan J. Nutri., 3(3): 154-157.
- Stobbs TH (1971). Production and composition of milk from cows grazing siratro (Phaseolus atropurpureus) and greenleaf desmodium (*Desmodium intortus*). Aust. J. Exp. Agric. Anim. Husbandry, 11: 268-273.
- Tibayungwa F, Mugisha JYT, Nabasirye M (2009). Dairy heifer growth and time to mating weight when fed elephant grass as sole feed: A simulation model. In: Chilliard Y, Glasser F, Faulconnier Y, Bocquier F, Veissier I, Doreau M (eds.) Ruminant physiology. Digestion, metabolism, and effects of nutrition on reproduction and welfare. Wageningen Academic Publishers, Wageningen, The Netherlands, pp. 782-783.
- Twinamasiko NI (2001). Dairy production. In: Mukiibi JK (ed) Agriculture in Uganda. Vol. IV: Livestock and Fisheries. 404 pp. National Agricultural Research Organization-CTA-Fountain Publishers, Kampala, Uganda, pp. 18-42.
- Webster, AJF (1993). The metabolizable protein system for ruminants. In: Haresign W, Garnsworthy PC (eds) Recent advances in animal nutrition - 1992, pp. 93-110. Butterworths, London, UK..